# PHOTOSYNTHESIS BY ISOLATED CHLOROPLASTS

# V. PHOSPHORYLATION AND CARBON DIOXIDE FIXATION BY BROKEN CHLOROPLASTS

by

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The attainment of complete extracellular photosynthesis by chloroplasts has made it possible to study photosynthesis as an isolated process divorced from the structural and metabolic complexity of a whole  $\operatorname{cell^{1,\,2}}$ . It has now been found that extracellular photosynthesis does not depend on the structural integrity of chloroplasts. This paper reports experiments in which whole chloroplasts were broken and active photosynthetic systems reconstituted by a recombination of chloroplast fractions and several cofactors. With proper additions, the photosynthetic activity of the reconstituted system, as measured by  $\operatorname{CO}_2$  fixation per unit of chlorophyll, was several times greater than that of the intact chloroplasts.

The fractionation of chloroplasts and the reconstitution of photosynthetically active systems with broken chloroplasts provides further experimental support for the previously discussed concept<sup>1, 2, 3</sup> that whole chloroplasts are complete cytoplasmic structures, specialized for photosynthesis, which contain three groups of enzymes, each group catalyzing an increasingly complex phase of the process: photolysis of water, photosynthetic phosphorylation, and CO<sub>2</sub> fixation.

### **METHODS**

Preparation of broken chloroplasts (P1w) by treatment with water

Whole chloroplasts  $(P_1)$  were prepared by grinding 100 g spinach leaves, freshly harvested from the greenhouse, in 0.35 M NaCl as described previously<sup>1</sup>. After removal of sand and large cell debris the whole chloroplasts were collected from the green suspension by centrifugation for 7 minutes at 1000 g at 0° C. The chloroplasts were washed once with 0.35 M NaCl and collected again by centrifugation. The washed whole chloroplasts were then suspended in about 50 ml of ice-cold water. The water treatment caused the chloroplasts to become swollen and irregular in shape; they could no longer be sedimented at 1000 g. The water-treated chloroplasts are referred to in this paper as "broken chloroplasts"  $(P_{1w})$ .

The chlorophyll content of each chloroplast suspension was determined and an aliquot con-

taining 0.5 mg chlorophyll was used in each reaction vessel.

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# Preparation of chloroplast extract (CE)

Whole chloroplasts were prepared from 300 g spinach leaves as described above. The leaves were ground in 3 batches, each of 100 g and the washed chloroplasts from all three were suspended in water to give a total volume of 30 ml. A small aliquot of the suspension was taken for chlorophyll estimation 1 and the remaining suspension centrifuged at 20,000 g for 10 minutes at 0° C. The green sediment was discarded and the clear, straw-colored supernatant liquid was decanted and designated "chloroplast extract" (CE). It was found convenient to express the concentration of the chloroplast extract on the basis of the chlorophyll in the chloroplasts from which the extract originated.

In some cases the chloroplast extract was lyophilized and stored as a dry powder at  $-18^{\circ}$  C. Aqueous solutions of this material were used. Here the quantity of the lyophilized powder used was calculated from the chlorophyll content of the chloroplasts from which it was derived. Even when stored at  $-18^{\circ}$  C the lyophilized material lost about 50% of its activity in 7-14 days.

## Measurement of photochemical reactions with broken chloroplasts

Photolysis, photosynthetic phosphorylation, and  $CO_2$  fixation were measured as described previously except that the reaction mixtures were made to volume by the addition of glass-distilled water instead of the 0.35 M NaCl formerly used with whole chloroplasts<sup>1</sup>.

### RESULTS

# Photosynthetic phosphorylation with broken chloroplasts

In our earlier experiments<sup>1,3,4</sup> active photosynthetic phosphorylation was obtained only with whole chloroplasts. It has now been found that on destroying the structure of the whole chloroplasts by treatment with water their capacity for photosynthetic phosphorylation was unimpaired when the requisite cofactors,  $Mg^{++}$ ,  $FMN^*$ , vitamin  $K_3$  (2-methyl-1,4-naphthoquinone), and ascorbate, were added under anaerobic conditions. As shown in Table I, the response of broken chloroplasts to the addition of these cofactors was similar to that of whole chloroplasts<sup>5,6</sup>.

Photosynthetic phosphorylation by broken chloroplasts depended on the addition of soluble cofactors only. The enzymes involved in photosynthetic phosphorylation were apparently unaffected by the disruption of chloroplast structure by the water treatment, and remained bound to the broken chloroplast particles.

TABLE I

COMPARISON OF EFFECTS OF COFACTORS ON PHOTOSYNTHETIC PHOSPHORYLATION
BY WHOLE AND BROKEN CHLOROPLASTS

(See the preceding paper<sup>1</sup> for experimental details)

Additions 4 6 1	(µ <b>m</b> oles)		Pi esterifu	red (µmoles)	
Vitamin K <sub>2</sub>	FMN	и	Vhole chloroplasts	Broken chloroplasts	
o	0.1	Complete	7.0	10.5	
0.003	0.1	Complete	6.8	9.5	
0.03	0.01	Complete	11.7	11.5	
0.03	· O. I	Complete	14.3	17.2	
0.3	0	Complete	16.0	18.2	
0.03	0.1	Complete	12.6	20.0	
0.03	0.1	Mg omitted	0.34	5.7	
0.03	0.1	Ascorbate omitt	ed 2.7	1.9	

<sup>\*</sup> The following abbreviations will be used in this paper: ADP, ATP, adenosine di- and tri-phosphate; FMN, flavin mononucleotide; DPN, TPN, di- and tri-phosphopyridine nucleotides.

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# CO<sub>2</sub> fixation by broken chloroplasts

The ability of chloroplasts to fix  $CO_2^7$  was almost completely lost when their structure was damaged by treatment with water. Their  $CO_2$ -fixing capacity was restored upon the addition of the clear, straw-colored water extract (CE) from whole chloroplasts (Table II). The addition of CE to the broken chloroplasts  $(P_{1w})$  gave a small increase in  $CO_2$  fixation in the dark;  $CO_2$  fixation in the light was approximately 10 times as great as that in the dark, leaving little doubt that the addition of CE influenced principally the light-dependent  $CO_2$  fixation.

TABLE II  ${\it EFFECT~OF~CHLOROPLAST~EXTRACT~(CE)} \ \ {\it on~photosynthetic~and~dark~carbon~dioxide~fixation~by~broken~chloroplasts~(P_{1W})$ 

(See text and	preceding	paper1	for o	experimental details	)

	14CO <sub>2</sub> fix	red (c.p.m.)			
Expt. No.	$P_{1w}$	$P_{1w} + CE$	P <sub>1w</sub> + CE Dark fixation		
54	2700	108,300	14,700		
56	1950	149,400	7,200		
65	1900	276,800	24,400		
67	950	137,300			

Table III shows that the addition to the broken chloroplasts of DPN, TPN, or ATP, singly or in combination, failed to restore their capacity for  $CO_2$  fixation in the absence of CE, but significantly increased photosynthetic  $CO_2$  fixation in the presence of CE. Heating the CE almost completely abolished its effect on  $CO_2$  fixation by broken chloroplasts.

TABLE III

effect of adenosine triphosphate (ATP) and di- and triphosphopyridine nucleotides (DPN and TPN) on the fixation of  $^{14}\mathrm{CO}_2$  by broken chloroplasts ( $P_{1\mathrm{w}}$ ) supplemented with a water extract of chloroplasts (CE)

(See text and preceding paper<sup>1</sup> for experimental details)

4.4492		14CO <sub>2</sub> fixed (c.p.m.)	
Additions	$P_{1w}$	$P_{1\omega} + CE$	P <sub>1w</sub> + heated CE
None	950	137,300	19,750
ATP	1300	204,950	35,400
DPN	2250	347,700	20,250
TPN	1600	450,250	19,450
ATP + DPN	2000	411,950	35,550
ATP + TPN	2800	659,350	29,900

The stimulating effect of pyridine nucleotides on  $CO_2$  fixation by broken chloroplasts supplemented with CE was of special interest. No such effect of pyridine nucleotides was observed on photosynthetic phosphorylation with either whole or broken chloroplasts, nor, as reported earlier<sup>3</sup>, on  $CO_2$  fixation by whole chloroplasts. References p. 468.

Fig. 1 indicates that the effect of pyridine nucleotides on  $\mathrm{CO}_2$  fixation by broken chloroplasts varied with the concentration of CE in the reaction mixtures. At high

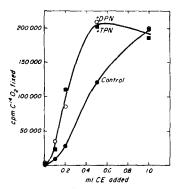


Fig. 1. Effect of diphosphopyridine nucleotide (DPN) and triphosphopyridine nucleotide (TPN) on CO<sub>2</sub> fixation by broken chloroplasts supplemented with varying amounts of chloroplast extract (CE). Experimental conditions as given in text.

concentrations of CE the effect of added pyridine nucleotides disappeared. It seems reasonable to conclude that pyridine nucleotides which appear to be required for  $CO_2$  fixation, though not for photosynthetic phosphorylation, are present in adequate amounts in whole chloroplasts. Only when these cofactors are leached out by the water treatment of the whole chloroplasts could a dependence of  $CO_2$  fixation on pyridine nucleotides be experimentally demonstrated. The effect of added TPN was, on an equimolar basis, somewhat greater than that of DPN. The significance of this observation must await future work.

The reconstituted broken chloroplast system reduced CO<sub>2</sub> to the level of carbohydrates. The products, shown in Fig. 2, were similar to those obtained with whole chloroplasts<sup>7</sup> and included phosphate esters of fructose, glucose, ribulose, dihydroxy acetone, and glyceric acid; glycine, alanine,

glycolic acid, glucose and dihydroxyacetone. The reconstituted system differed from whole chloroplasts in that appreciable quantities of starch were not synthesized unless soluble starch was added.

The photosynthetic activity of the reconstituted broken chloroplast system indicates that treating whole chloroplasts with water removes both soluble enzymes and cofactors, such as ATP and pyridine nucleotides, which are required for CO<sub>2</sub> fixation. The identification of the enzymes contained in CE is now in progress; among those already identified are the glyceraldehyde phosphate dehydrogenases of green leaves8, carboxydismutase9, phosphoglyceryl kinase, phosphopentokinase, phosphoglucomutase, and phosphorylase. The role of these and other water-soluble enzymes of chloroplasts in CO<sub>2</sub> fixation is currently under investigation.

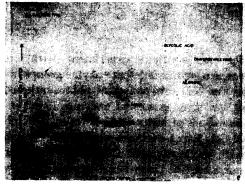
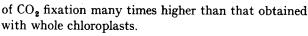


Fig. 2. Chromatogram showing products of photosynthetic CO<sub>2</sub> fixation by broken chloroplasts. Compound directly above area labeled "monophosphates" has been identified as phosphoglyceric acid. Experimental procedures as given in text.

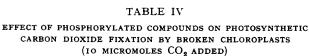
Photosynthetic CO<sub>2</sub> fixation by broken chloroplasts supplemented with chloroplast extract, TPN, and ATP equalled or exceeded that obtained with the most active preparations of whole chloroplasts. A further several-fold increase in the rate of CO<sub>2</sub> fixation was obtained by the addition of any one of a number of compounds, principally phosphorylated sugars (Table IV). The products of CO<sub>2</sub> fixation were again similar to those obtained with whole chloroplasts. As seen from the diagrammatic

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summary in Fig. 3, the combined effect of the various addenda has resulted in a level



The stimulating effect of manganese on  $CO_2$  fixation previously noted with whole chloroplasts<sup>7</sup> was much more pronounced with broken chloroplasts supplemented with CE and all the cofactors described.  $CO_2$  fixation was reduced approximately eighty per cent when manganese was omitted.



Additions	CO <sub>2</sub> fixed, µmoles
None*	2.0
3-Phosphoglyceric acid	3.7
Ribose-5-phosphate	4.0
Glucose-1-phosphate	4.4
Glucose-6-phosphate	3.8
Fructose-6-phosphate	3.1
Fructose diphosphate	5.2

<sup>\*</sup>  $P_{1w}$ , CE, ATP, TPN.

3 µmoles of each phosphorylated compound was added to the reaction mixture. Other experimental conditions as described in text and in the preceding paper.

# P. CE. AT P, T PN P. CE, AT P, T PN

Fig. 3. Diagrammatic representation of the effects of various addenda on  $\mathrm{CO_3}$  fixation by broken chloroplasts. Experimental conditions as in text. Abbreviations:  $P_1$ , whole chloroplasts;  $P_{\mathrm{IW}}$ , water-treated chloroplasts; CE, chloroplast extract; ATP, adenosine triphosphate; DPN, TPN, di- and triphosphopyridine nucleotides; FDP, fructose-1, 6-diphosphate.

### DISCUSSION

The photosynthetic experiments with broken chloroplasts provide further evidence concerning the different enzyme groups in whole chloroplasts¹ and their participation in the different phases of photosynthesis. Broken chloroplasts contained only two of the three groups of enzymes found in whole chloroplasts: those controlling the photolysis of water and photosynthetic phosphorylation. At least some of the group of CO<sub>2</sub>-fixing enzymes were leached out by treating whole chloroplasts with water, with the result that CO<sub>2</sub> fixation was completely abolished. The phosphorylating enzymes, like the enzymes of photolysis, were not water-soluble and remained bound to the particles.

A distinction should be made between broken chloroplasts  $(P_{1w})$  described here and the chloroplast fragments  $(P_2)$  described earlier. The  $P_{1w}$  preparation was obtained from whole chloroplasts first separated from other cytoplasmic particles and then broken, not by mechanical means, but by treatment with water. The  $P_2$  preparation consisted of a mixture of chloroplasts, mechanically fragmented in the grinding of leaves, with other small cytoplasmic particles, which probably included mitochondria. The feeble capacity for photosynthetic phosphorylation of the  $P_2$  preparation, as contrasted with the  $P_{1w}$ , may have resulted from damage during the mechanical breaking of the chloroplasts and also from possible interference by the non-chloroplast particles.

The technique of breaking whole chloroplasts by treating them with water has References p. 468.

provided a direct experimental demonstration of the cofactors required for CO<sub>2</sub> fixation. The effect of ATP in restoring the capacity for CO<sub>2</sub> fixation to broken chloroplasts is in harmony with the conclusion<sup>1,2</sup> that ATP is essential for CO<sub>2</sub> fixation. As with whole chloroplasts, phosphorylated sugars, known to be formed with the aid of ATP, were identified among the products of CO<sub>2</sub> fixation by broken chloroplasts (Fig. 2).

It seems likely that pyridine nucleotides (PN+) serve as electron carriers in the reduction of CO<sub>2</sub>. The reduction of pyridine nucleotides, linked to photolysis of water, could be aided by ATP (compare<sup>10-13</sup>) as represented by Equation 1. The reduced pyridine nucleotide (PNH), again with the aid of ATP, would then reduce CO<sub>2</sub> in accordance with Equation 2:

$$Light + H2O + PN+ + ATP \rightarrow PNH + H+ + \frac{1}{2}O2 + ADP + Pi$$
 (I)

$$CO_2 + 2PNH + 2H^+ + nATP \longrightarrow (CH_2O) + 2PN^+ + nADP + nP_i$$
 (2)

The inclusion of ATP in reaction I is attractive for it permits us to retain the concept

discussed elsewhere<sup>2</sup> that the key reaction in photosynthesis, the photolysis of water, is accomplished by the energy of I quantum of red light (ca. 43 kcal). The pyrophosphate bonds of ATP could provide the supplementary energy for chloroplast reactions which require an input of more than 43 kcal, such as reduction of pyridine nucleotides. Taking the standard oxidation-reduction potential of TPN at pH 7 as—0.32 V<sup>14</sup>, it would require approximately 50 kcal to give a TPN<sub>red</sub>/TPN<sub>ox</sub> ratio of I.

The reduction of CO<sub>2</sub> to the level of carbohydrate (Equation 2) almost certainly involves a reduction of a carboxyl to a carbonyl group. The

LIGHT

(0) — H<sub>2</sub>0

cytochromes?

ascorbate

vit, K

FMN

2 {H}

CO<sub>2</sub>

sugar phosphates

STARCH

Fig. 4. Scheme for photosynthesis by isolated chloroplasts. Photolysis of water (center) leading either to ATP synthesis and the reconstitution of water (right) or to CO<sub>2</sub> reduction (below) linked to oxygen evolution (left).

reducing potential of pyridine nucleotides is insufficient to accomplish this step without a lowering of the potential barrier, which again could be accomplished by phosphorylation of the carboxyl group by ATP<sup>15</sup>.

The overall scheme for photosynthesis by chloroplasts<sup>1</sup> has now been modified (Fig. 4) to include pyridine nucleotides in the CO<sub>2</sub> reduction pathway.

# SUMMARY

1. Photosynthetic esterification of inorganic phosphate into adenosine triphosphate, and reduction of CO<sub>2</sub> to the level of carbohydrate, hitherto found to occur only in whole chloroplasts, have now been observed with chloroplasts broken by treatment with water.

2. Broken chloroplasts retained only two of the three groups of enzymes contained in whole chloroplasts, namely, those controlling the photolysis of water and photosynthetic phosphorylation. At least some of the enzymes concerned in reduction of CO<sub>2</sub> were leached out by treating the chloroplasts with water, with the result that CO<sub>2</sub> fixation was completely abolished.

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- On addition of the requisite cofactors, the capacity of broken chloroplasts for photosynthetic phosphorylation was the same as that of whole chloroplasts.
- 4. The restoration to broken chloroplasts of the full capacity for photosynthetic CO<sub>2</sub> fixation of whole chloroplasts required the addition of pyridine nucleotides, adenosine triphosphate, and the soluble enzymes removed by water treatment of whole chloroplasts.
- 5. An additional several-fold increase in the rate of CO<sub>2</sub> fixation by the reconstituted broken chloroplast system was obtained by the further addition of one of several compounds, principally phosphorylated sugars. This has resulted in a level of CO<sub>2</sub> fixation by broken chloroplasts which is much higher than that obtained with whole chloroplasts.

### REFERENCES

- 1 D. I. ARNON, M. B. ALLEN AND F. R. WHATLEY, Biochim. Biophys. Acta, 20 (1956) 449.
- <sup>2</sup> D. I. Arnon, Science, 122 (1955) 9.
- <sup>3</sup> D. I. ARNON, M. N. ALLEN AND F. R. WHATLEY, Nature, 174 (1954) 394-
- <sup>4</sup> D. I. Arnon, F. R. Whatley and M. B. Allen, J. Am. Chem. Soc., 76 (1954) 6324.
- <sup>5</sup> F. R. Whatley, M. B. Allen and D. I. Arnon, Biochim. Biophys. Acta, 16 (1955) 605.
- <sup>6</sup> D. I. Arnon, F. R. Whatley and M. B. Allen, ibid., 16 (1955) 607.
- <sup>7</sup> M. B. Allen, D. I. Arnon, J. B. Capindale, F. R. Whatley and L. J. Durham, J. Am. Chem. Soc., 77 (1955) 4149.
- <sup>8</sup> L. L. Rosenberg and D. I. Arnon, J. Biol. Chem., 217 (1955) 361.
- 9 R. C. Fuller, F. R. Whatley, M. B. Allen and D. I. Arnon (unpublished data).
- 10 O. KANDLER, Z. Naturforsch., 5b (1950) 423.
- 11 B. L. Strehler, Phosphorus Metabolism, II (1952) 491.
- 12 C. E. SLATER, Ann. Rev. Biochem., 22 (1953) 17.
- 13 J. S. C. WESSELS, Rec. trav. chim., 73 (1954) 529.
- 14 K. Burton and T. H. Wilson, Biochem. J., 54 (1953) 86.
- 15 F. LIPMANN, Advances in Enzymol., I (1941) 99.

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